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Moran GJ, Amii RN, Abrahamian FM, Talan DA (2005). Methicillinresistant *Staphylococcus aureus* in community-acquired skin infections. *Emerg. Infect. Dis.* 11: 928-930.

Pitout JDD, Church DL, Gregson DB, Chow BL, McCracken M, Mulvey M, Laupland KB (2007). Molecular epidemiology of CTXM-producing *Escherichia coli* in the Calgary Health Region: emergence of CTX-M-15-producing isolates. *Antimicrob. Agents Chemother.* 51: 1281-1286.

Pelczar JR, Harley JP, Klein DA (1993). *Microbiology: Concepts and Applications.* McGraw-Hill Inc., New York, pp. 591-603.

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Full Length Research Paper

The codon 17 polymorphism of the *CTLA4* gene in type 1 diabetes mellitus in the Baghdad population

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Accepted 13 January, 2014

The aim of this work was to study the polymorphism in *CTLA4* gene in insulin-dependent diabetes mellitus (IDDM) type I patients in Baghdad population. To achieve this goal, blood samples were collected from 80 IDDM (40 males and 40 females) and 20 samples of healthy, DNA was isolated and the *CTLA4* gene (A 152 bp fragment) were amplified by using specific primers for exon1 of this gene, and then found the sequence of this region. The DNA sequencing results of flank sense of *CTLA4* gene from healthy patients was found to be compatible (100%) with wild type of *Homo sapiens* from the Gene Bank, while 99% compatibility were found for the gene from 70 IDDM cases patients with wild type of gene. The difference may be attributed to one transition mutations, A/G at position 49 of the *CTLA4* gene (from AGC to AAC). It is a missense mutation that leads to changes in amino acid from serine (S) to asparagine (N). Our results showed that the incidence of A/G mutation at nucleotide position 49 and diabetes was highly significant ($X^2 = 100$, $P < 0.01$). In total, 12% of patients with IDDM (10 cases) had two transition mutation +49 A/G and +47 C/T single nucleotide polymorphism from total cases, 98% compatibility were found for that gene from 10 IDDM cases patients with wild type of gene. The +47 C/T SNP was silent mutation which resulted in change of codon from GGT to GGC but no changes translated to amino acid (glycine to glycine). However, there was no significant correlation between diabetes and incidence of C/T at nucleotide 47 ($X^2 = 0.055$, $P > 0.05$). In conclusion, our case study suggests that the +49 A/G SNP of the *CTLA4* gene is strongly associated with genetic susceptibility to type 1 diabetes mellitus in the Baghdad/Iraqi population.

Key words: *CTLA4* gene, insulin-dependent diabetes mellitus, A/G polymorphism.

INTRODUCTION

Cytotoxic T-lymphocyte antigen 4 (*CTLA4*) also known as cluster of differentiation 152 (CD152) is a protein that plays an important role in the immune system regulations. *CTLA4* is a member of the immunoglobulin superfamily, which is expressed on the surface of Helper T cells and transmits an inhibitory signal to T cells. The *CTLA4* encodes the T cell receptor involved in the control of T cell proliferation and mediates T cell apoptosis (Yanagawa et al., 1997; Larsen et al., 1999). The receptor protein is a specific T lymphocyte surface

antigen that is detected on cells only after antigen presentation. Thus, *CTLA4* is directly involved in both immune and autoimmune responses and may be involved in the pathogenesis of multiple T cell-mediated autoimmune disorders. The human *CTLA4* gene is located at chromosome 2q33 (Nistico et al., 1996; Donner et al., 1997a). This gene is a member of the immunoglobulin superfamily and encodes a protein which transmits an inhibitory signal to T cells. The protein contains a V domain of 116 amino acids, a

transmembrane domain, and a cytoplasmic tail. Alternate transcriptional splice variants, encoding different isoforms, have been characterized. The membrane-bound isoform functions as a homodimer which is interconnected by a disulfide bond, while the soluble isoform functions as a monomer (Kristiansen et al., 2000).

An A-to-G substitution at nucleotide 49 in exon 1 results in an amino acid substitution (Thr/Ala) in the leader peptide of the protein (Donner et al., 1997b). The Ala allele has been shown to predispose the individual carrying it to the development of various immune diseases including insulin-dependent diabetes mellitus, Graves disease, Hashimoto thyroiditis, celiac disease, systemic lupus erythematosus, thyroid-associated orbitopathy, and other autoimmune diseases (Anjos and Polychronakos, 2004).

Mutations and polymorphisms in this gene results in alteration of the *CTLA4* activity and are believed to play an important role in the risk of developing autoimmunity (Anjos and Polychronakos, 2004). The *CTLA4* (49+) GG homozygous genotype is associated with Type 1 diabetes in Egyptian children especially with younger age of onset and in younger patients and not associated with grades of diabetic control or diabetic complication (Hatem et al., 2008; Mosaad et al., 2012). The aim of this study was to assess the contribution of this *CTLA4* polymorphism to the susceptibility to type 1 diabetes in the Baghdad population.

MATERIALS AND METHODS

Samples and DNA extraction

Whole blood samples were obtained from 80 Baghdad patients affected by insulin-dependent diabetes mellitus (IDDM) (40 males and 40 females, age ranged from 4 to 25 years). Samples from 20 healthy individuals were used as a control group. In total, 4 ml whole blood was collected into an Ethylenediaminetetraacetic acid (EDTA) tube. The samples were stored at -20°C until further processing. DNA was extracted by DNA extraction kit (Wizard® Genomic DNA Purification Kit, Promega, Madison, WI, USA) according to the manufacturer's protocol.

Amplification of exon 1 of *CTLA4* gene

A 152 bp fragment containing the +49 A/G polymorphism in exon 1 of *CTLA4* was amplified using a forward primer (*CTLA4*: 5'-AAGGCTCAGCTGAACCTGGT-3') and a reverse primer (*CTLA4*: 5'-CTGCTGAAACAAATGAAACCC-3') (Alpha DNA Company, Canada) (Marron et al., 1997). The polymerase chain reaction (PCR) amplification was performed in a total volume of 25 µl containing 5 µl DNA, 12.5 µl Go Taq green master mix 2X (Promega corporation, USA), 1 µl of each primer (50 pmol). The thermal cycling conditions were as follows: Denaturation at 94°C for 5 min, followed by 30 cycles of 94°C for 30 s, 57°C for 1 min and 72°C for 1 min with final incubation at 72°C for 7 min using a thermal Cycler (Gene Amp, PCR system 9700; Applied Biosystem) (Genc et al., 2004; Hatem et al., 2008). The PCR products were separated by 1.5% agarose gel electrophoresis, stained with ethidium bromide and visualized by ultraviolet light (302 nm).

Sequencing and sequence alignment

Sequencing of exon 1 of *CTLA4* gene was performed by Macro gen company, USA. Homology search was conducted using Basic local alignment search tool (BLAST) program which is available at the National Center Biotechnology Information (NCBI) online at (<http://www.ncbi.nlm.nih.gov>) and BioEdit program. The results were compared with data obtained from Gene Bank published ExPASY program which is available at the NCBI online.

Statistical analysis

The statistical analysis is a very important final step in the research to analyses and evaluates the obtained results. Medical statistics of this study was conducted via computer based statistical program which was: X² for Windows computer package. The statistical analysis tests used in this were as follows: P value < 0.01 is considered a significant correlation.

RESULTS AND DISCUSSION

CTLA4 gene was successfully amplified using specific PCR primers for exon 1. Figure 1 showed PCR amplification of exon 1 of the *CTLA4* where a specific product at 152 bp was observed. Our result is in agreement with other studies (Hatem et al., 2008; Waterhouse et al., 1995). Sequencing of this gene was performed to detect variant +49A/G which related to development of diabetes. Sequences alignment using BLAST and BioEdit showed the 100% similarity or homology of healthy sample with wild type of the *CTLA4* gene of *H. sapiens* from the Gene Bank (Figure 2). The *CTLA4* gene from 70 diabetes patients shows 99% compatibility with the wild type sequences of *CTLA4* gene from Gene Bank as shown in Figure 3A, there are one transition at position +49 A/G single nucleotide polymorphism that cause a serine to asparagine substitution in codon 17, there is a high significance between diabetes and incidence of + 49 A/G position in exon 1 of *CTLA4* gene (X² = 100, P > 0.01), Table 1 shows the type of mutation and the effect of these mutations and Table 2 shows the translation of *CTLA4* gene of all groups (healthy and patient) to a protein sequence, and two transition mutation at position +47 C/T and +49 A/G of *CTLA4* gene from 10 diabetes patients was identified.

The sequence shows 98% compatibility with wild type *CTLA4* gene as shown in Figure 3B. single nucleotide polymorphism at position 47+ C/T that silent mutation, no change translate amino acid (Glycine to Glycine), there is lower significant correlation between type 1 diabetes and incidence of this SNP, (X² = 0.055, P > 0.05). Most molecular epidemiology studies have evaluated the role of the +49A/G single nucleotide polymorphism that causes a threonine to alanine substitution in codon 17 and associated with altered protein expression (Anjos et al., 2002) and T-cell activation (Maurer et al., 2002). Gribben et al. (1995) have suggested that this may be through antigen specific induction of the apoptotic pathway. The mentioned study investigated the A49G polymorphism in

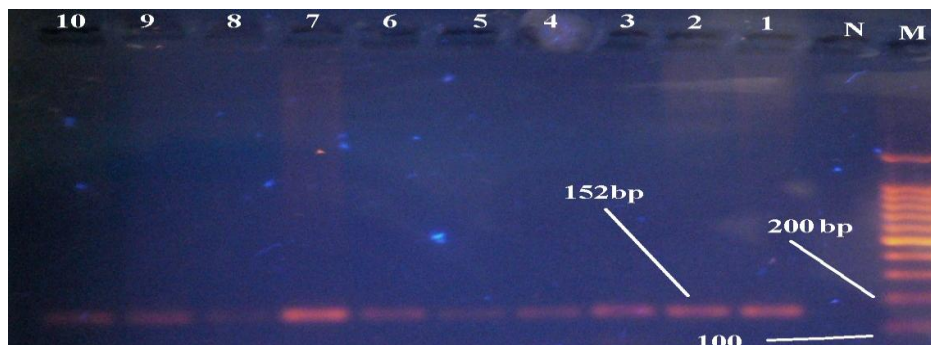


Figure 1. Agarose gel electrophoresis for detection of amplified *CTLA4* gene. Bands were fractionated by electrophoresis on a 1.5 % agarose gel (2 h., 5 V/cm, 1× Tris-acetic buffer) and visualized under UV light after staining with ethidium bromide. Lane: 12 (M:100 bp ladder); Lane: N (negative control); Lane: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 (PCR product).

Homo sapiens chromosome 2 genomic contig, features in this part of subject sequence:

[cytotoxic T-lymphocyte protein 4](#)

Score = 150 bits (81), Expect = 2e-34, Identities = 81/81 (100%), Gaps = 0/81 (0%)

Query 16 CTTTGCAGAAGACAGGGATGAAGAGAAGAAAAACAGGAGAGTGCAGGGCCAGGTCCTGG 75

|||||

Sbjct 264 CTTTGCAGAAGACAGGGATGAAGAGAAGAAAAACAGGAGAGTGCAGGGCCAGGTCCTGG 205

Query 76 TAGCCAGGTTTCAGCTGAGCCT 96

|||||

Sbjct 204 TAGCCAGGTTTCAGCTGAGCCT 184

Figure 2. Sequencing of sense flanking the partial *CTLA4* gene for healthy as compared with wild type *CTLA4* obtained from Gene Bank.

exon 1 of *CTLA4* gene in 40 Lebanese and 46 controls from the same ethnic background.

An increase in the frequency of the G allele was discovered in patients when compared to control subjects, this difference was statistically significant, despite the small sample size. Wafai et al. (2011) showed an association of *CTLA4* with type 1 diabetes in Lebanese population. An association was detected between the *CTLA4* gene polymorphism and younger-onset type 1 diabetes with autoimmune thyroid disease (AITD) (Gough, et al., 2005).. The G variant was suggested to be genetically linked to AITD-associated type 1 diabetes of younger onset in this Japanese population (Mochizuk, et al., 2003). The defect in these patients presumably lies in a T-cell mediated autoimmune mechanism (Takara et al., 2000). Chistiakov et al. (2001) reported that the *CTLA4* gene is strongly associated with insulin-dependent diabetes mellitus (IDDM) in a fifty-six families each consisting of two siblings (one affected with IDDM diagnosed before the age of 18 years and one non-diabetic sibling).

It was reported that the *CTLA4* 49 (A/G) mutation

conferred a risk of type 1 diabetes in the Chinese children but not in the West African children. On the other hand, the novel *CTLA4* 159 (C/G) mutation conferred a risk of type 1 diabetes in the West African children but not in the Chinese type 1 diabetic children (Hyiaman et al., 2001). Donner et al. (1997b) showed that an alanine at codon 17 of *CTLA4* is associated with genetic susceptibility to Graves disease as well as to IDDM. Lemos et al. (2009) states that the *CTLA4* +49 A/G polymorphism is not associated with susceptibility to type 1 diabetes mellitus in the Portuguese population. This contrasts with positive associations that have been reported for the +49A/G polymorphism in case control studies in populations from Belgium, Germany, Poland, France, Japan, China, Italy, the Philippines, Lebanon, Estonia and Iran (Zalloua et al., 2004; Kavvoura and Ioannidis, 2005; Mojtahedi et al., 2005). However, lack of association for the + 49A/G polymorphism has also been reported in populations from the USA, Japan, Ghana, UK, France, Czech Republic, Morocco, Argentina, Brazil and Azerbaijan (Marron et al., 1997; Caputo et al., 2005; Hauache et al., 2005; Kavvoura

A: Sense of the partial *CTLA4* gene, shown one transition mutation.

Score = 172 bits (93), Expect = 4e-41 , Identities = 95/96 (99%), Gaps = 0/96 (0%)

Query 1 AAAAGTCTCACTCACCTTTGCAGAAGACAGGGATGAAGAGAAGAAAAACAGGAGAGTGC 60
 |||
 Sbjct 54942207 AAAAGTCTCACTCACCTTTGCAGAAGACAGGGATGAAGAGAAGAAAAACAGGAGAGTGC
 54942148

Query 61 AGGGCCAGGTCCTGGTAA^ACCAGGTTTCAGCTGAGCCT 96
 |||
 Sbjct 54942147 AGGGCCAGGTCCTGGTAA^GCCAGGTTTCAGCTGAGCCT 54942112

B: Sense of the partial *CTLA4* gene, shown two transition mutation.

Homo sapiens chromosome 2 genomic contig, features in this part of subject sequence:
 cytotoxic T-lymphocyte protein 4
 Score = 165 bits (89), Expect = 7e-39, Identities = 93/95 (98%), Gaps = 0/95 (0%)

Query 1 AAAGTCTCACTCACCTTTGCAGAAGACAGGGATGAAGAGAAGAAAAACAGGAGAGTGCA 60
 |||
 Sbjct 54942206 AAAGTCTCACTCACCTTTGCAGAAGACAGGGATGAAGAGAAGAAAAACAGGAGAGTGCA
 54942147

Query 61 GGGCCAGGTCCTGGCA^ACCAGGTTTCAGCTGAGCCT 95
 |||
 Sbjct 54942146 GGGCCAGGTCCTGGT^AAGCCAGGTTTCAGCTGAGCCT 54942112

Figure 3. Sequencing of sense flanking the *CTLA4* gene for diabetes as compared with wild type *CTLA4* obtained from Gene Bank , (A: 70 diabetes patients have one mutation ; B: 10 diabetes patients have two mutation)

Table 1. Types of mutations detected in partial *CTLA4* gene of diabetes patients.

No.	Location of gene bank	Nucleotide change	No. of sample	Amino acid change	Predicted effect	Type of mutation
1	A/G 49+	AGC > AAC	70	Serine (S)/ Asparagine (N)	Missense	Transition
2	C/T 47+	GGT > GGC	10	Glycine (G) / Glycine (G)	Silent	Single nucleotide polymorphism

Table 2. Amino acid sequences of healthy and patient group.

No.	Sequencing of amino acid	Sample
1	KVSLTFAEDRDEEKKKQESAGPGPGSQVQLSLK	20 healthy
2	KVSLTFAEDRDEEKKKQESAGPGPGNQVQLSLK	70 Patient
3	KVSLTFAEDRDEEKKKQESAGPGPGNQVQLSLK	10 patient

K: lysine; V: Valine; S: Serine; L: Lysine; T: Threonine; F: Phenylalanine; A: Alanine; E: Glutamic acid; D: Asparagine; R: Arginine; Q: Glutamine; G: Glycine; P: Proline.

and Ioannidis, 2005; Ahmedov et al., 2006).

Conclusion

Our study showed that there was significant correlation

between diabetes and incidence of A/G +49 position in exon 1 of *CTLA4* gene, despite the limited size of our sample, our results together with population studies show an association of *CTLA4* with type 1 diabetes mellitus, on the other hand, the novel of +47 C/T silent mutation was no significant correlation between type 1 diabetes in

Baghdad population.

REFERENCES

- Ahmedov G, Ahmedova L, Sedlakova P, Cinek O (2006). Genetic association of type 1 diabetes in an Azerbaijanian population: the HLA-DQ-DRB1*04, the insulin gene, and CTLA4. *Pediatr. Diabetes* 7:88.
- Anjos S, Polychronakos C (2004). Mechanism of genetic susceptibility to type I diabetes: beyond HLA. *Mol. Genet. Metab.* 81(3):187-195.
- Chistiakov D, Savost'anov K, Nosikov V (2001). CTLA4 gene polymorphisms are associated with and linked to insulin-dependent diabetes mellitus in a Russian population. *BMC Genet.* 2:6.
- Donner H, Braun J, Seidl C, Rau H, Finke R, Ventz M, Walfish PG, Usadel KH, Badenhoop K (1997a). Codon 17 polymorphism of the cytotoxic T lymphocyte antigen 4 gene in Hashimoto's thyroiditis and Addison's disease. *J. Clin. Endocrinol. Metab.* 82(12):4130-4132.
- Donner H, Rau H, Walfish PG, Braun J, Siegmund T, Finke R, Herwig JU, Usadel K, Badenhoop K (1997b). CTLA4 Alanine-17 Confers Genetic Susceptibility to Graves' Disease and to Type 1 Diabetes Mellitus. *J. Clin. Endocrinol. Metab.* 82:143-146.
- Genc S, Genc K, Sercan O (2004). Analysis of cytotoxic T lymphocyte antigen-4 (CTLA-4) exon 1 polymorphism in patients with type 1 diabetes mellitus in a Turkish population. *J. Pediatr. Endocrinol. Metab.* 17:731-735.
- Gough SC, Walker LS, Sansom DM (2005). CTLA4 gene polymorphism and autoimmunity. *Immunol. Rev.* 204:102.
- Gribben JG, Freeman GJ, Boussiotis VA (1995). CTLA4 mediates antigen specific apoptosis of human T cells. *Proc. Natl. Acad. Sci.* 92:811-815.
- Hauache OM, Reis AF, Oliveira CS, Vieira JG, Sjorros M, Ilonen J (2005). Estimation of diabetes risk in Brazilian population by typing for polymorphisms in HLA-DR-DQ, INS and CTLA-4 genes. *Dis. Markers.* 21:139.
- Hatem MS, Nestor R, Michael L (2008). The CTLA4-819 C/T and +49 A/G dimorphisms are associated with Type1 diabetes in Egyptian children. *Indian J. Hum. Genet.* 14(3):92-98.
- Hyiaman D, Hou L, Zhiyin R, Zhiming Z, Yu H, Amankwah A, Harada S (2001). Association of a novel point mutation (C159 G) of the *CTLA4* gene with type 1 diabetes in West Africans but not in Chinese. *Diabetes* 50:2169-2171.
- Kavvoura FK, Ioannidis JP (2005). CTLA-4 gene polymorphisms and susceptibility to type 1 diabetes mellitus: a HuGE review and meta-analysis. *Am. J. Epidemiol.* 162:3.
- Kristiansen OP, Larsen ZM, Pociot F (2000). CTLA4 in autoimmune disease a general susceptibility gene to autoimmunity. *Genes Immunol.* 1:170-184.
- Larsen ZM, Kristiansen OP, Mato E (1999). IDDM12 (CTLA4) on 2q33 and IDDM13 on 2q34 in genetic susceptibility to type 1 diabetes (insulin-dependent). *J. Autoimmun.* 31(1):35-42.
- Lemos M, Coutinho E, Gomes L, Bastos M, Fagulha A, Barros L, Carrilho F, Geraldés E, Regateiro F, Carvalheiro M (2009). The *CTLA4*+ 49 A/G polymorphism is not associated with susceptibility to type 1 diabetes mellitus in the Portuguese population. *Int. J. Immunogenet.* 36:193-195.
- Marron MP, Raffel LJ, Garchon HJ, Jacob CO, Serrano-Rios M, Martinez-Larrad MT, Teng VP, Park J, Zhang ZX, Goldstein DR, Tao JW, Beaurain G, Bach JF, Huang HS, Luo DF, Zeidler A, Rotter JI, Yang MC, Modilevsky T, Maclaren NK, She JX (1997). Insulin-dependent diabetes mellitus (IDDM) is associated with CTLA4 polymorphisms in multiple ethnic groups. *Hum. Mol. Genet.* 6(8):1275-1282.
- Mochizuk M, Amemiya S, Kobayashi K, Shimura Y, Ishihara T, Nakagomi Y, Onigata K, Tamai S, Kasuga A, Nanazawa S (2003). Association of the CTLA-4 Gene 49 A/G Polymorphism with Type 1 Diabetes and Autoimmune Thyroid Disease in Japanese Children. *Diabetes Care* 26(3):843-847.
- Mojtahedi Z, Omrani GR, Doroudchi M, Ghaderi A (2005). CTLA-4 +49A/G polymorphism is associated with predisposition to type 1 diabetes in Iranians. *Diabetes Res. Clin. Pract.* 68:111.
- Mosaad Y, Elsharkawy A, El-Deek B (2012). Association of CTLA4 (+49A/G) gene polymorphism with type 1 diabetes mellitus in Egyptian Children. *Immunol. Investig.* 41:28-37.
- Nistico L, Buzzetti R, Pritchard LE (1996). The CTLA-4 gene region of chromosome 2q33 is linked to and associated with, type 1 diabetes. *Belgian Diabetes Registry. Hum. Mol. Genet.* 5:1075-1080.
- Takara M, Komiya I, Kinjo Y, Tomoyose T, Yamahiro S, Akamine H, Masuda M, Takasu N (2000). Association of CTLA-4 Gene A/G Polymorphism in Japanese Type 1 Diabetic Patients With Younger Age of Onset and Autoimmune Thyroid Disease. *Diabetes Care.* 23:975-978.
- Waterhouse P, Penninger JM, Timms E, Wakeham A, Stahinaian A, Lee KP, Thompson CB, Griesser H, Mak TW (1995). Lymphoproliferative disorders with early lethality in mice deficient in *Ctla-4*. *Sci.* 270:985-988.
- Wafai R, Chmairie H, Makki R, Fakhour H (2011). Association of HLA class II alleles and CTLA4 polymorphism with Type 1 diabetes. *Saudi J. Kidney Dis. Transplant* 22(2):273-281.
- Yanagawa T, Taniyama M, Enomoto S, Gomi K, Maruyama H, Ban Y, Saruta T (1997). CTLA4 gene polymorphism confers genetic susceptibility to Graves' disease in Japanese. *Thyroid* 7:843-846.
- Zalloua PA, Abchee A, Shbaklo H, Zreik TG, Terwedow H, Halaby G, Azar ST (2004). Patients with early onset of type 1 diabetes have significantly higher GG genotype at position 49 of the CTLA4 gene. *Hum. Immunol.* 65:71.

Full Length Research Paper

Mutation N308T of protein tyrosine phosphatase SHP-2 in two Senegalese patients with Noonan syndrome

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Noonan syndrome is a genetic autosomal dominant disorder characterized by facial dysmorphism, short stature, delayed puberty and congenital heart defects. The first gene implicated in this syndrome is PTPN11, encoding protein tyrosine phosphatase SHP-2. Several studies worldwide have identified missense mutations in this gene in patients with Noonan syndrome. Our objective focused on mutations screening of PTPN11 on a Senegalese population with Noonan syndrome. Six patients clinically diagnosed with Noonan syndrome were included in this study. DNA was extracted from whole blood by phenol chloroform. Mutation screening was performed by bidirectional sequencing of amplified polymerase chain reaction (PCR) products of PTPN11 exons frequently mutated in Noonan syndrome. This study identified in two patients, a c.923A>C mutation in exon 8, predicting Asn308Thr (N308T) on SHP-2 protein. This is the first time that this mutation is described in Noonan syndrome in Africa, while codon 308 was reported as a hot spot mutation site in other populations. Frequently reported amino acid substitutions were Asn308Asp and Asn308Ser. All these mutations affected the protein tyrosine phosphatase domain (PTP) of SHP-2 protein exerting a gain of function which would likely explain observed phenotypes in patients.

Key words: Mutation, N308T, protein tyrosine phosphatase (PTP), SHP-2 protein, Noonan syndrome, Senegal.

INTRODUCTION

Noonan syndrome (NS, MIM 163950) is an autosomal dominant dysmorphic syndrome described first by Noonan (1968). The prevalence of NS is estimated to be 1 in 1000 to 2500 births. The disease is characterized by proportionate short stature, delayed puberty, congenital heart defects and multiple minor anomalies such as hypertelorism, malrotated ears, webbed neck, bleeding diathesis, cryptorchidism in males, mental retardation, and hearing difficulties (Marino et al., 1999; Roberts et

al., 2013; van der Burgt et al., 2007). The most common congenital heart defect is pulmonary valve stenosis with displastic leaflets followed by hypertrophic obstructive cardiomyopathy (HC), atrial septal defects (Marino et al., 1999; Musante et al., 2003; Tartaglia et al., 2006).

Jamieson et al. (1994) mapped a gene for NS to the long arm of chromosome 12 (12q24). Tartaglia et al. (2001) reported that NS is caused by heterozygous missense mutations of the gene protein-tyrosinephosph-

atase nonreceptor type 11 (PTPN11) located on 12q24. PTPN11 mutations were detected in 45% of unrelated individuals with sporadic or familial NS (Tartaglia et al., 2002). PTPN11 gene consists of 15 exons and the expressed protein encodes cytoplasmic tyrosine phosphatase with two tandemly arranged Src homology 2 (SH2) domains (N-SH2 and C-SH2) at the N terminal region and a C terminal protein-tyrosine phosphatase domain (PTP) (Chan and Feng, 2007). The PTPN11 gene is widely expressed in various human tissues, especially in the heart, brain, and skeletal muscle (Ahmad et al., 1993). The protein plays a critical role in regulating the response of eukaryotic cells to extracellular signals through the RAS/MAPK pathway (Roberts et al., 2013).

All PTPN11 missense mutations associated with NS were clustered in the interacting portions of the N-SH2 domain and the PTP domains are involved in switching the protein from its inactive to the active conformation. Functional studies by energetic-based structural analysis of two N-SH2 mutants revealed that those mutations favoured the active conformation of PTPN11 protein, resulting in a gain-of-function effect (Tartaglia et al., 2001; Uhlen et al., 2006). Also, other studies have reported enhanced phosphatase activity of NS mutants located in the SH2 and PTP domains (Niihori et al., 2005; Tartaglia et al., 2003).

Understanding of the molecular genetic causes of NS, enable the study of the pathophysiological mechanisms underlying the varied medical and developmental features of NS. PTPN11 belongs to the RAS-MAPK pathway which is an important signal transduction pathway. Mutations that cause NS deregulated this pathway leading to the clinical features observed. Furthermore, all the other genes implicated in NS including SOS1, RAF1, and KRAS encode proteins integral to this pathway (Roberts et al., 2013).

Several studies have reported mutation analysis of PTPN11 gene and genotype-phenotype correlation in NS in different geographical regions (Bertola et al., 2004; Pierpont et al., 2009; Sznajder et al., 2007; Tartaglia et al., 2006; Yoshida et al., 2004). Mutation screening in NS patients from United States showed that all mutations are exonic changes with the majority clustering in exon 3 and 8. The most common mutation was a c.922A>G in exon 8, leading to the Asn308Asp substitution within the PTP domain (Tartaglia et al., 2002). The occurrence of an adjacent c.923A>G mutation predicting an Asn308Ser change and c.923A>C (Asn308Thr) indicated that codon 308 is a hot spot site for NS with a frequency of 36% in PTPN11 mutated patients (Tartaglia et al., 2002; Tartaglia et al., 2006; Tartaglia et al., 2003). Other studies have reported the occurrence of different mutations types of PTPN11 in European and Asian populations. In Germany, the most common mutation is c.188A>G (Tyr63Cys) followed by c.922A>G (Musante et al., 2003), while in Japan the most common mutation is c.236A>G

(Gln79Arg) (Yoshida et al., 2004). In Africa, few studies have focused on mutation screening in NS. Only one report from Morocco identified the c.922A>G mutation in two affected siblings with normal parents (Elalaoui et al., 2010) and c.182A>G (Ratbi et al., 2008). In sub-Saharan Africa, no report from PTPN11 mutation is available to date. The objective in this study was to screen for PTPN11 gene mutations in 6 Senegalese patients with NS and summarized observed clinical features.

POPULATION AND METHODS

Patients and clinical assessment

After informed consent, 6 patients clinically diagnosed with NS were included in this study. Patients were examined by clinicians from the Cardiology Unit of Fann Hospital in Dakar (Senegal), who have experience with NS. Electrocardiograms, echocardiograms and clinical photographs were obtained from each patient. NS was diagnosed on the basis of the presence of the following major characteristics: typical facial dysmorphism, pulmonary valve stenosis or hypertrophic cardiomyopathy, chest deformity, developmental delay and cryptorchidism in male patients. To be diagnosed for NS, patient with facial dysmorphism had to have at least two of the major characteristics (van der Burgt et al., 1994).

Mutation screening of PTPN11 gene

Genomic DNA was isolated from peripheral blood lymphocytes by classic phenol/chloroform method. The most frequently mutated exons reported in NS (3, 4, 7 and 8) and flanking introns of PTPN11 gene were amplified by PCR with sets of primers as described previously (Tartaglia et al., 2002). All PCR products were purified with QIAquick PCR purification kit from Qiagen™ as described by the manufacturer. Mutations were screened by direct bidirectional sequencing of the purified PCR products with Big Dye Terminator chemistry (Perkin Elmer Biosystem™) on an ABI 3100 auto sequencer (ABI™, Foster City, CA). Obtained sequences were analysed by BioEdit software (Hall, 1997).

RESULTS AND DISCUSSION

Clinical features of studied NS patients are summarized in Table 1. In this study, 4 females and 2 males were recruited with ages ranging from 1 to 31 years. None of the patients had known family history of NS. Growth was delayed in all cases. Dysmorphic features were present with variable severity in all cases. Figure 1 illustrates some of these features (hypertelorism, low-set ears, webbed neck, chest deformity, and ptosis) in a male and a female patient. The most frequent cardiac abnormalities were hypertrophic cardiomyopathy (HCM) and pulmonary valve stenosis (PS). One of the male cases had cryptorchidism.

The phenotypes observed in NS are heterogeneous and vary in different ages. The diagnosis of Noonan syndrome is primarily clinical and is guided by the most common dysmorphic signs such as hypertelorism, low-

Table 1. Clinical features in 6 Senegalese patients with NS.

Parameter	NS1	NS2	NS3	NS4	NS5	NS6
Age (years)	17	31	4	15	1	4
Sex	F	F	M	M	F	F
Growth	delayed	delayed	delayed	delayed	delayed	delayed
Craniofacial dysmorphism	Hypertelorism webbed neck	Hypertelorism	Hypertelorism webbed neck ptosis	Hypertelorism	Hypertelorism	Hypertelorism webbed neck low set ears
Chest	Deformity	-	Deformity	Deformity	-	Deformity
Cardiovascular defects	HC	-	HC	PS	HC	PS
Genital defects	-	-	Cryptorchidism	-	-	-

NS: Noonan syndrome; HC: hypertrophic cardiomyopathy; PS: pulmonary valve stenosis.

Table 2. PTPN11 codon 308 mutations in Senegalese patients with NS compared to other populations around the world.

Exon 8	rs number	AA change	Senegal	USA	USA	Brazil	Japan
			Ndiaye et al	Tartaglia et al. (2006)	Pierpont et al. (2009)	Bertola et al. (2006)	Yoshida et al. (2004)
c. 922A>G	rs28933386	N308D	-	40/204	4/33	-	2/18
c. 923A>G	rs121918455	N308S	-	13/204	2/33	1/21	-
c. 923A>C	ND	N308T	2/6	2/204	1/33	-	-

ND: Not determined.

set ears, chest deformities and short stature, associated cardiac abnormalities (Tartaglia et al., 2002; van der Burgt, 2007; van der Burgt et al., 1994). These morphological abnormalities were observed in most of our patients and were the key elements of diagnosis.

In addition to these morphological abnormalities, alterations in several genes have been implicated in NS. This is the case of the PTPN11 gene which is mutated in 40 to 50% of patients with NS (Tartaglia et al., 2001). Mutation screening of PTPN11 gene in studied exons have identified a heterozygote substitution in exon 8, c.923A>C, in two unrelated patients NS5 and NS6 (Figure 2). This missense mutation led to N308T substitution on the PTP domain of SHP-2 protein.

This mutation has not been detected in 15 healthy Senegalese controls.

The mutation rate found in our study (2 of the 6 studied patients) is lower than reported in other populations (Hung et al., 2007; Tartaglia et al., 2002). This difference could be explained, first by the small number of patients enrolled, due to the absence of a clinical consultation in dysmorphology at health facilities in Senegal. The six patients recruited are indeed followed up in the Cardiology Unit of Aristide Le Dantec Hospital for their associated heart abnormalities. Secondly, this study only have sequenced the exons 3, 4, 7 and 8 which are most frequently mutated in NS but mutations may be observed in other exons not studied (Musante et al., 2003; Yoshida et al.,

2004). Thirdly, PTPN11 is not the only gene associated with NS. Mutations have also been reported in genes such as KRAS, RAF, SOS1 all involved in the same signaling RAS/MAP pathway (Roberts et al., 2013).

The c.923A>C mutation observed in our study is heterozygous and confirms the autosomal dominant transmission of NS. This mutation leads to the substitution of N308T in SHP-2 protein. It is the first time that this mutation is reported in African patients with NS. Two previous studies have reported this mutation in USA in 3 individuals without any details about their Caucasian or African-American ethnic origin (Pierpont et al., 2009; Tartaglia et al., 2006) (Table 2). It was hypothesized that N308T as de novo mutation may



Figure 1. Two NS patients: (A and B) a 4 year old boy (NS3) without mutations in studied exons of PTPN11 gene, typical signs of dysmorphism are ptosis, hypertelorism, webbed neck, chest deformity; (C and D) a 4 year old girl (NS6) with mutation c.923A>C in exon 8 of PTPN11 gene, typical signs of dysmorphism are low set ears, hypertelorism, webbed neck.

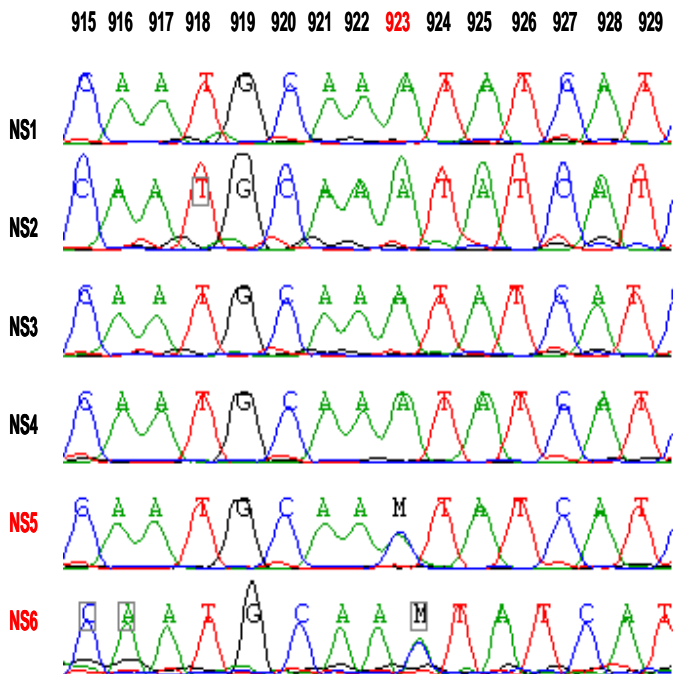


Figure 2. Sequence plots alignment around position 923 of exon 8 of PTPN11 gene in studied patients. NS1 to NS4 are homozygous for the wild type allele at position 923 (genotype A/A); NS5 and NS6 have a heterozygous mutation c.923A>C (genotype A/C).

arise first in Africa. It may be confirmed by investigating the ethnic origin of US patients bearing this mutation.

Although, position 308 of SHP-2 protein have been reported to be the most frequently mutated site in NS, amino-acid substitution reported were mostly N308D and N308S (Keren, 2006; Musante et al., 2003; Tartaglia et al., 2001). These mutations are located in the PTP domain of SHP-2. Crystallographic data have shown that the PTP domain as the SH2 domain play an important role in the stability and function of the SHP-2 protein (Musante et al., 2003; Tartaglia et al., 2001). Amino-acid changes in interaction sites between PTP and N-SH2 domains or near these sites may lead to the switch of SHP-2 protein from its inactive form to an active form, which result in a dominant positive effect on the activation of the RAS/MAPK signaling pathway effect. This activation is responsible for the morphological and cardiac anomalies in Noonan syndrome as reported in mouse model (Chen et al., 2010; Lapinski et al., 2013). Although inactivation of this pathway can rescue congenital heart defects and craniofacial malformations in Noonan mouse model (Nakamura et al., 2007, 2009).

It is also established that N308D mutation lead to a milder hyperactivation of the RAS/MAPK pathway compared to other described mutations in the PTPN11 gene (Oishi et al., 2006). Similarly, patients with Noonan syndrome and carrying this mutation have normal psychomotor development (Sznajder, 2009). This is not the case with the mutated patients in our study (NS5, NS6), which showed growth delay. This could be explained by the difference of involved amino acids. Indeed, it would be appropriate to consider functional studies on mutation N308T in order to evaluate its effect on the SHP-2 protein activity.

Conclusions

This study focused on finding the PTPN11 gene mutations involved in Senegalese patients with Noonan syndrome. Two patients in six had a heterozygous mutation, c.923A>C in exon 8 of PTPN11 gene, resulting in amino acid change Asn308Thr in SHP-2 protein. This is the first time that this mutation is described in Noonan syndrome in Africa although position 308 is considered as a "hot spot" site. The results presented are preliminary results of a pilot study of the PTPN11 gene in Senegalese patients with Noonan syndrome and currently followed in the health services in Dakar.

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ABBREVIATIONS

NS, Noonan syndrome; **PTPN11**, protein tyrosine phosphatase non receptor 11; **PTP**, protein tyrosine phosphatase domain; **HC**, hypertrophic cardiomyopathy; **PS**, pulmonary valve stenosis.

REFERENCES

- Ahmad S, Banville D, Zhao Z, Fischer EH, Shen SH (1993). A widely expressed human protein-tyrosine phosphatase containing src homology 2 domains. *Proc. Natl. Acad. Sci. U S A* 90:2197-201.
- Bertola DR, Pereira AC, de Oliveira PS, Kim CA, Krieger JE (2004). Clinical variability in a Noonan syndrome family with a new PTPN11 gene mutation. *Am. J. Med. Genet. A* 130A:378-83.
- Chan RJ, Feng GS (2007). PTPN11 is the first identified proto-oncogene that encodes a tyrosine phosphatase. *Blood* 109:862-7.
- Chen PC, Wakimoto H, Conner D, Araki T, Yuan T, Roberts A, Seidman C, Bronson R, Neel B, Seidman JG, Kucherlapati R (2010). Activation of multiple signaling pathways causes developmental defects in mice with a Noonan syndrome-associated *Sos1* mutation. *J. Clin. Invest.* 120:4353-65.
- Elalaoui SC, Kraoua L, Liger C, Ratbi I, Cave H, Sefiani A (2010). Germinal mosaicism in Noonan syndrome: A family with two affected siblings of normal parents. *Am. J. Med. Genet. A*. 152A:2850-3.
- Hall T (1997). BioEdit Software. In Biosciences (ed.): An Abbott company. Carlsbad.
- Hung CS, Lin JL, Lee YJ, Lin SP, Chao MC, Lo FS (2007). Mutational analysis of PTPN11 gene in Taiwanese children with Noonan syndrome. *J. Formos. Med. Assoc.* 106:169-72.
- Jamieson CR, van der Burgt I, Brady AF, Van Reen M, Elswawi MM, Hol F, Jeffery S, Patton MA, Mariman E (1994). Mapping a gene for Noonan syndrome to the long arm of chromosome 12. *Nat. Genet.* 8:357-60.
- Keren B (2006). Syndrome de Noonan et mutations du gène PTPN11 : corrélation génotype-phénotype, Thèse Université PARIS VAL-DE-MARNE Faculté de Médecine de Créteil, Paris.
- Lapinski PE, Meyer MF, Feng GS, Kamiya N, King PD (2013). Deletion of SHP-2 in mesenchymal stem cells causes growth retardation, limb and chest deformity, and calvarial defects in mice. *Dis. Model. Mech.* 6:1448-58.
- Marino B, Digilio MC, Toscano A, Giannotti A, Dallapiccola B (1999). Congenital heart diseases in children with Noonan syndrome: An expanded cardiac spectrum with high prevalence of atrioventricular canal. *J. Pediatr.* 135:703-6.
- Musante L, Kehl HG, Majewski F, Meinecke P, Schweiger S, Gillissen-Kaesbach G, Wieczorek D, Hinkel GK, Tinschert S, Hoeltzenbein M, Ropers HH, Kalscheuer VM (2003). Spectrum of mutations in PTPN11 and genotype-phenotype correlation in 96 patients with Noonan syndrome and five patients with cardio-facio-cutaneous syndrome. *Eur. J. Hum. Genet.* 11:201-6.
- Nakamura T, Colbert M, Krenz M, Molkenin JD, Hahn HS, Dorn GW, 2nd, Robbins J (2007). Mediating ERK 1/2 signaling rescues congenital heart defects in a mouse model of Noonan syndrome. *J. Clin. Invest.* 117:2123-32.
- Nakamura T, Gulick J, Pratt R, Robbins J (2009). Noonan syndrome is associated with enhanced pERK activity, the repression of which can prevent craniofacial malformations. *Proc. Natl. Acad. Sci. USA.* 106:15436-41.
- Niihori T, Aoki Y, Ohashi H, Kurosawa K, Kondoh T, Ishikiriyama S, Kawame H, Kamasaki H, Yamanaka T, Takada F, Nishio K, Sakurai M, Tamai H, Nagashima T, Suzuki Y, Kure S, Fujii K, Imaizumi M, Matsubara Y (2005). Functional analysis of PTPN11/SHP-2 mutants identified in Noonan syndrome and childhood leukemia. *J. Hum. Genet.* 50:192-202.
- Noonan JA (1968). Hypertelorism with Turner phenotype. A new syndrome with associated congenital heart disease. *Am. J. Dis. Child.* 116:373-80.
- Oishi K, Gaengel K, Krishnamoorthy S, Kamiya K, Kim IK, Ying H, Weber U, Perkins LA, Tartaglia M, Mlodzik M, Pick L, Gelb BD (2006). Transgenic Drosophila models of Noonan syndrome causing PTPN11 gain-of-function mutations. *Hum. Mol. Genet.* 15:543-53.
- Pierpont EI, Pierpont ME, Mendelsohn NJ, Roberts AE, Tworog-Dube E, Seidenberg MS (2009). Genotype differences in cognitive functioning in Noonan syndrome. *Genes. Brain. Behav.* 8:275-82.
- Ratbi I, Gati AE, Sefiani A (2008). The Moroccan human mutation database. *Indian. J. Hum. Genet.* 14:106-7.
- Roberts AE, Allanson JE, Tartaglia M, Gelb BD (2013). Noonan syndrome. *Lancet* 381:333-42.
- Sznajder Y (2009). Etude des manifestations cardiovasculaires chez les patients présentant un syndrome de Noonan porteurs de mutation au sein du gène PTPN11; rôles des gènes de la voie de signalisation des MAP kinases pour les syndromes apparentés, Thèse Université Libre de Bruxelles, Bruxelles.
- Sznajder Y, Keren B, Baumann C, Pereira S, Alberti C, Elion J, Cave H, Verloes A (2007). The spectrum of cardiac anomalies in Noonan syndrome as a result of mutations in the PTPN11 gene *Pediatrics* 119:e1325-31.
- Tartaglia M, Kalidas K, Shaw A, Song X, Musat DL, Van der Burgt I, Brunner HG, Bertola DR, Crosby A, Ion A, Kucherlapati RS, Jeffery S, Patton MA, Gelb BD (2002). PTPN11 mutations in Noonan syndrome: molecular spectrum, genotype-phenotype correlation, and phenotypic heterogeneity. *Am. J. Hum. Genet.* 70:1555-63.
- Tartaglia M, Martinelli S, Stella L, Bocchinfuso G, Flex E, Cordeddu V, Zampino G, Burgt I, Palleschi A, Petrucci TC, Sorcini M, Schoch C, Foa R, Emanuel PD, Gelb BD (2006). Diversity and functional consequences of germline and somatic PTPN11 mutations in human disease. *Am. J. Hum. Genet.* 78:279-90.
- Tartaglia M, Mehler EL, Goldberg R, Zampino G, Brunner HG, Kremer H, Van der Burgt I, Crosby AH, Ion A, Jeffery S, Kalidas K, Patton MA, Kucherlapati RS, Gelb BD (2001). Mutations in PTPN11, encoding the protein tyrosine phosphatase SHP-2, cause Noonan syndrome. *Nat. Genet.* 29:465-8.
- Tartaglia M, Niemeyer CM, Fragale A, Song X, Buechner J, Jung A, Hahlen K, Hasle H, Licht JD, and Gelb BD (2003). Somatic mutations in PTPN11 in juvenile myelomonocytic leukemia, myelodysplastic syndromes and acute myeloid leukemia. *Nat. Genet.* 34:148-50.
- Uhlen P, Burch PM, Zito CI, Estrada M, Ehrlich BE, Bennett AM (2006). Gain-of-function/Noonan syndrome SHP-2/Ptpn11 mutants enhance calcium oscillations and impair NFAT signaling. *Proc. Natl. Acad. Sci. USA.* 103:2160-5.
- van der Burgt I (2007). Noonan syndrome. *Orphanet J. Rare Dis.* 2:4. Not cited
- van der Burgt I, Berends E, Lommen E, van Beersum S, Hamel B, Mariman E (1994). Clinical and molecular studies in a large Dutch family with Noonan syndrome. *Am. J. Med. Genet.* 53:187-91.
- Van der Burgt I, Kupsky W, Stassou S, Nadroo A, Barroso C, Diem A, Kratz CP, Dvorsky R, Ahmadian MR, Zenker M (2007). Myopathy caused by HRAS germline mutations: implications for disturbed myogenic differentiation in the presence of constitutive HRas activation. *J. Med. Genet.* 44:459-62.
- Yoshida R, Hasegawa T, Hasegawa Y, Nagai T, Kinoshita E, Tanaka Y, Kanegane H, Ohyama K, Onishi T, Hanew K, Okuyama T, Horikawa R, Tanaka T, Ogata T (2004). Protein-tyrosine phosphatase, nonreceptor type 11 mutation analysis and clinical assessment in 45 patients with Noonan syndrome. *J. Clin. Endocrinol. Metab.* 89:3359-64

Full Length Research Paper

A study of correlation between CYP2C9 gene polymorphism and Warfarin maintenance dose in anticoagulant therapy among Han people in Yunnan of China

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In this study, we investigated the correlation between CYP2C9 gene polymorphism and maintenance dose of Warfarin in 300 patients who were the Han population derived from the Affiliated Yan An Hospital of Kunming Medical University in Yunnan Province of China, subjected to the operation of cardiac valve replacement and Warfarin oral administration post operation from 2008 to 2009 by detecting the genotypes and Allele Frequency at the three candidate loci (CYP2C9*2, CYP2C9*3 and CYP2C9*c_65) of CYP2C9 gene from the blood samples. Polymerase chain reaction -restriction fragment length polymorphism (PCR-RFLP) assay and DNA sequencing were used to ascertain the genotypes and their corresponding distribution rate. The maintenance dose of Warfarin administered in anticoagulant therapy among the CYP2C9*3 genotypes found in our experiment showed: A/A wild type > A/C heterozygote > C/C homozygote, suggesting that patients with C mutation need the lowest maintenance dose of Warfarin among the three genotypes found in this study. Our results will shed a new light on the personalized medicine of Warfarin and provide basic and genetic experimental data and foundation for future studies with regard to multiple genes' effect on Warfarin dosage in anticoagulant therapy.

Key words: Warfarin, maintenance dosage, CYP2C9, gene polymorphism, single nucleotide polymorphism (SNP), restriction fragment length polymorphism (RFLP), correlation.

INTRODUCTION

As a kind of common oral anticoagulant, Warfarin is extensively applied for the anticoagulant therapy in various diseases, including valvular heart disease or pathological conditions, valve replacement, fibrillation atrial, electrical conversion, coronary heart disease, pulmonary embolism, deep vein thrombosis and stroke e.t.c. Along with

the elevation of incidence rate in chronic fibrillation atrial, cardiovascular and cerebrovascular diseases related to thrombo embolism occur more than ever before. Additionally, the popularity of artificial cardiac valve replacement results in more and more patients receiving long-term oral administration of Warfarin for anticoagulant

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therapy. However, in the clinical practice of anticoagulant therapy, it is difficult to control the effective maintenance dose of Warfarin, due to the large variation with regard to intra-individual difference in maintenance dose of anticoagulant agent, its therapeutic effect and side effect (Aithal et al., 1999).

Inadaptable dose of Warfarin administration, especially over dose may lead to some serious complications including hemorrhage or thrombosis, even a threat to life. In the early stage of Warfarin administration, the incidence rate of hemorrhage is about 12%, and death rate resulted from hemorrhage in patients is 2% (Levine et al., 2001). It is reported that in America, there are about 2 million patients who were suffered from the side effect of drugs, half of which were lethal (Lazarou et al., 1998). Therefore, to make a change for traditional medication mode into personalized medicine will shed a new light on the settlement of this difficulty by adjusting the Warfarin dose as optimal one suitable for patients and, at the same time, reducing the side effect of it.

To date, among the studies of gene polymorphism of CYP2C9, the most studied and extensive ones were involving in the correlation between mutation of either CYP2C9*2 or CYP2C9*3 and clinical Warfarin maintenance dosage (Higashi et al., 2002). Furthermore, there exists large variation in Allele Frequency of CYP2C9*2 and CYP2C9*3 among different races. However, until now, there are few reports about the roles of gene loci of CYP2C9 in Warfarin administration, and the guidance of Warfarin personalized medication needs intensive study.

In this study, we investigated the correlation between the gene polymorphism of three important gene loci (CYP2C9*2, CYP2C9*3 and CYP2C9*c₆₅) in CYP2C9 gene and Warfarin maintenance dosage administered in anticoagulant therapy, so as to pave a new way for the personalized medicine of Warfarin according to different genotypes of different patients.

MATERIALS AND METHODS

Object of study

Sample harvesting and Admission standard of patients

A total of 300 patients of Han population who were subjected to cardiac valve replacement from 2008 to 2009 in Affiliated Yan An Hospital of Kunming Medical University in Yunnan Province of China were registered and recruited according to the strict standard (see below) in this study. After informed consent was signed, a total of 3ml peripheral venous blood was extracted from each patient. The blood samples were anticoagulated by addition of ethylenediaminetetraacetic acid (EDTA) and preserved at -48°C.

All 300 patients, more than 18 year-old, were orally administered with Warfarin for anticoagulant therapy under strict monitor persisted for one month post of operation, whose international normalized ratio (INR) range from 1.5 to 3.0. In the retrospective whole therapies of the patients were performed at this Hospital. The Warfarin tablets administered to all these patients were produced from the same pharmaceuticals company (Orion Corporation). The

strict monitor index consisted of normal hepatic function and obeying dietetic contraindication according to the medical order. Clinical therapeutic and laboratory data were recorded in detail.

Exclusion standard of samples

Under following circumstances, the patients were excluded outside of this study: The patients who had liver diseases at present and before, or his (her) serum transaminase was 1.5 fold more than that of normal level. Patients with impaired renal function, whose serum creatinine >120 μmol/L. In the prospective cases, Warfarin was orally administered three months prior to this test. Patients who were administered or administering the drugs influenced the metabolism of Warfarin Herman et al. (2006) Basal INR scale >1.4 - patients who were not appropriate for Warfarin administration due to other reasons.

Definition of target International normalized ratio (INR) scale and stable dosage of Warfarin

International normalized ratio (INR)

Since Professor Armand Quick (1935_a, 1935_b) set up routine prothrombin time (PT) blood coagulation assay in 1935, until now, it is still an important screen test to measure factors and associated inhibitors in exogenous blood coagulation system. However, the outcome of PT assay was influenced by various factors. Therefore, it must undergo standardization and quality control so as to elevate its precision, accuracy and reliability. In recent years, international normalized ratio (INR) detection was adopted extensively to measure the clotting time, which avoids the differential outcomes in different detections due to different reagents used in this assay.

Target International normalized ratio (INR) in this study

In this study, the scales of INR measured from blood samples of patients were screened by the standard ranging from 1.5 to 3.0, downward for 0.2 was considered as the normal. The patients whose INR were outside of this range were excluded from this study, because under this situation, the Warfarin dose could not attain stability, which is unfavorable to our study and may produce a misleading and even false outcome.

Stable dosages of Warfarin

Stable dosage of Warfarin was referred to: under the same dosage of Warfarin, sequential INR detection of patients ranged from 1.5 to 3.0. The interval of two INR tests was at least above 7 days.

Primer synthesis

The primers used in the PCR assay were synthesized by Sai Bai Sheng Gene Technology Co. Ltd. Primers of these 3 candidate gene loci were as follows:

CYP2C9*2: locates at 3rd exon

Single Nucleotide Polymorphism (SNP) name: rs1799853

Upriver primer 5'-ATGAAAACAGAGACTTACAGAGGT -3'

Downstream primer 5'-CCAGTAAGGTCAGTGATATGGAGTAG -3'

CYP2C9*3: locates at 7th exon SNP name: rs1057910

Upriver primer 5'-CTGAATTGCTACAACAATGTGCCA -3'

Downstream primer 5'-AGGCTGGTGGGGAGAAGGTCC -3'

CYP2C9*c₆₅: locates at 3rd exon SNP name: rs9332127

Upriver primer 5'-TTTTTGCTGTTAAGGGAATTTG -3'

Downstream primer 5'-CAATTCAGAGCTTGATCCATG -3'

Table S1. Contents and quantities of PCR reaction system.

Contents of PCR reaction system	Gene locus		
	CYP2C9*2 (quantity: µl)	CYP2C9*3 (quantity: µl)	CYP2C9*c_65 (quantity: µl)
Distilled water (PH8.2)	14	13	14
10×PCR Buffer	2.0	2.0	2.0
DNA template (250ng/µl)	1.0	2.0	1.0
dNTP	2.0	2.0	2.0
Upriver primer	0.5	0.5	0.5
Downstream primer	0.5	0.5	0.5
TaqDNA polymerase (5U/µl)	0.2	0.15	0.2

Table S2. Contents and quantities of the digestion system of CYP2C9*2, CYP2C9*3 and CYP2C9*c_65 gene locus.

Contents in Digestion system	Gene locus		
	CYP2C9*2 (Quantity: µl)	CYP2C9*3 (Quantity: µl)	CYP2C9*c_65 (Quantity: µl)
Distilled water (PH8.2)	7.7	5.7	6.7
10 × Buffer (Ava II)	0.8	0.8	0.8
Restrictive endonuclease (10u/ µl)	0.5	0.5	0.5
Products of PCR amplification	1.0	3.0	8.5

Selection of restrict endonuclease

CYP2C9*2: restrict enzyme of Ava II recognition site: GGTC*
 CYP2C9*3: restrict enzyme of Mva1 recognition site: CCA*GG
 CYP2C9*c_65: restrict enzyme of Hpa I recognition site: GTT*AAC

DNA extraction

Saturated phenol/ chloroform method was employed according to traditional procedure described previously by Joanna et al. (2012) to extract DNA from the blood samples of 300 patients who were recruited for this study.

DNA purification and concentration assaying were performed as described in Alessandra et al. (2011) report.

Primer dilution

The pairs of primers used in PCR amplification were diluted and adjusted at the concentration of 20 pmol/µl referred to following fomula:

$$\text{Volume of distilled water added } (\mu\text{l}) = \frac{2 \times \text{number of nmol of synthesized primer} \times 10^3}{40}$$

Following dilution, the primer solution was subpackaged and stored at -48°C, awaits further usage.

PCR amplification

PCR reaction systems are shown in Table S1. Pure water, 10×PCR reaction buffer, DNA template, dNTP, primers and Taq DNA polymerase was added into 0.2 ml sterile microcentrifuge tube

respectively and orderly, covered by paraffin oil. The tubes were placed into the PCR amplifier (GeneAmp PCR SYSTEM 9700, AB applied Biosystems) under conditions set up as following:

PCR reaction conditions of CYP2C9*2, CYP2C9*3 and CYP2C9*c_65 were shown below respectively:

97°C 5 min → 80°C 5 min → 94°C 1min → 60°C 1 min → 72°C 1 min → 72°C 10 min
 30 cycles
 97°C 5 min → 80°C 5 min → 94°C 30 s → 64.5°C 30 s → 72°C 30 s → 72°C 10 min
 30 cycles
 97°C 5 min → 80°C 5 min → 94°C 30 s → 55°C 30 s → 72°C 30 s → 72°C 10 min
 30 cycles

After PCR reaction was finished, the products obtained from PCR were taken out and stored at -20°C.

Digestion by nucleate endonuclease

Restrictive endonuclease Ava II, Mva1 and Hpa I were used for digestion in target fragment of CYP2C9*2, CYP2C9*3 and CYP2C9*c_65 gene loci respectively. The contents and quantities of the digestion system of CYP2C9*2, CYP2C9*3 and CYP2C9*c_65 gene loci were shown in Table S2. The digestion reaction was performed in a thermostatic waterbath at 37°C for 14 h.

Electrophoresis test for products from PCR amplification and digestion

In this test, 8% vertical native polyacrylamide gel electrophoresis

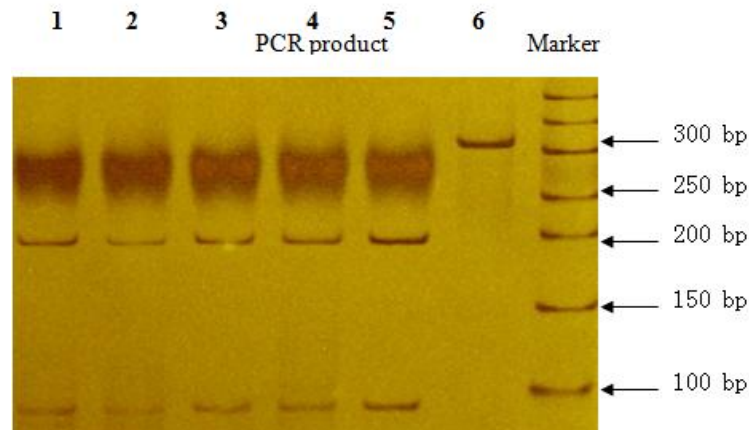


Figure 1. Samples 1 to 5 showed the digestive products following PCR, their genotypes were all C/C wild type. Sample 6 showed PCR product of CYP2C9*2 locus, whose segment size was 309 bp. Number 1 was heterozygote mutant A/C (127, 105, 75 and 22 bp), Number 2 was homozygote mutant C/C (105, 75 and 22 bp), Number 3 to 5 was wild type A/A (127 and 75 bp), Number 6 was the product of PCR amplification of CYP2C9*2 locus, with segment size of 202 bp.

was employed. Silver Nitrate staining was used to observe the outcomes of the electrophoresis. By using 50 and 100 bp DNA ladder as standard molecular weight markers, the lengths of DNA fragments of target genes were ascertained and their genotypes were determined. Silver Nitrate staining method was used for coloration. Camera (Canon, IXY DIGITAL) was used to take pictures of the staining outcome. Dry gelatin was made so as to preserve the outcome.

DNA sequencing

DNA sequencing of three loci of CYP2C9 gene following PCR amplification

The products of PCR amplification of CYP2C9*2 (rs1799853), CYP2C9*3 (rs1057910) and CYP2C9*c₆₅ (rs9332127) were subjected to DNA sequencing. A total of 5 DNA samples from each locus were used for DNA sequencing in order to ascertain whether the PCR amplification products were the expectant target fragments. 3130-Genetic Analysis Apparatus (ABI Company, America) was employed to perform automatic DNA sequencing.

DNA sequencing of three loci of CYP2C9 gene following PCR amplification and digestion

The digestive products underwent DNA sequencing at CYP2C9*3 (rs1057910) locus consisted of 5 cases of A/A wild type (127 bp, 75 bp), 1 case of C/C homozygote mutation type (105 bp, 75 bp, 22 bp) and 2 cases of A/C heterozygote mutant (127 bp, 105 bp, 75 bp, 22 bp).

The DNA sequencing for the digestive products of CYP2C9*c₆₅ (rs9332127) locus following PCR amplification included 5 cases of G/G wild type (317 and 54 bp) and 2 cases of G/C heterozygote mutant (371, 317 and 54 bp).

Statistical analysis

Experimental data were expressed as mean±SD and analyzed by

SPSS15.0 and PLINK software. Hardy-Weinberg law of genetic equilibrium was employed to detect the goodness of fit-test of genetic balance. The genotypes and allelotype frequency of three gene loci of CYP2C9 were calculated. χ^2 -test was used to analyze the data of genotypes and allelotype-frequency. A level of $p < 0.05$ was considered as statistical significance.

RESULTS

Products of PCR amplification and genotypes of three loci of CYP2C9 gene

The segment size of CYP2C9*2 locus derived from the PCR and digestion in 300 DNA samples was all 309 bp (Figure 1). With the digestion by restrictive endonuclease Ava II, when restriction enzyme cutting site was at base C, fragment of 309 bp-product following PCR was digested into three fragments, whose segment size was 195, 91 and 23 bp respectively. When restriction enzyme cutting site was at base T, the fragment of product following PCR could be digested into two fragments, whose segment size was 286 and 23 bp respectively. Only one allele- C (195, 91 and 23 bp) and one kind of genotype- C/C wild type (Figure 1) was checked out in the 300 samples. Among the 300 DNA samples, there was no mutational site was detected at CYP2C9*2 gene locus. As the digestive product of 23 bp fragment at CYP2C9*2 locus was the smallest one, when polyacrylamide gel electrophoresis (PAGE) finished, this fragment have run out of the gelatin, image of this fragment could not be observed within the gelatin. Only the 195 and 91 bp DNA fragments could be observed.

The segment size of CYP2C9*3 locus obtained from the PCR and digestion in 300 DNA samples was all 202

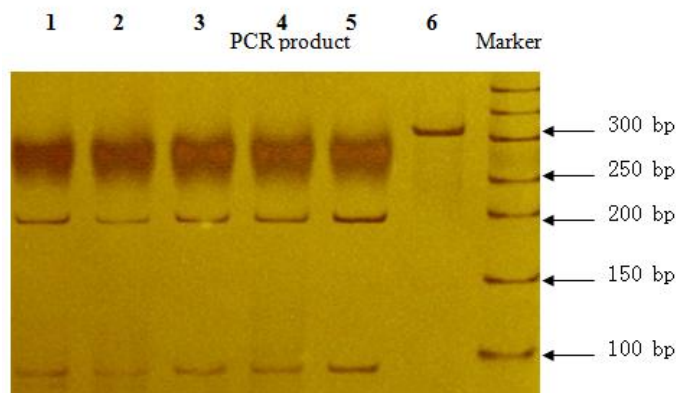


Figure S1. Samples 1 to 5 showed the digestive products following PCR, their genotypes were all C/C wild type. Sample 6 showed PCR product of CYP2C9*2 locus, whose segment size was 309 bp.

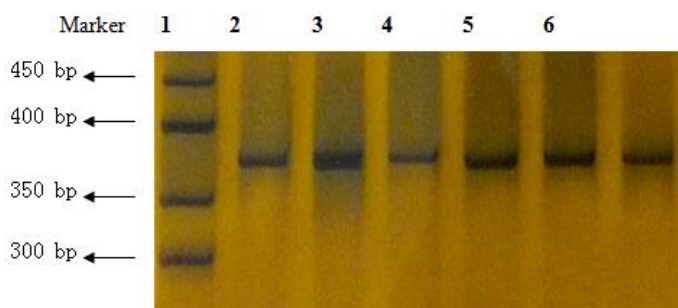


Figure S2. Number 1 to 6 showed PCR amplified fragments of CYP2C9*c_65 locus, with segment size of 371 bp.

bp (Figure S1). With restrictive nuclease *va1* digestion, when the restriction enzyme cutting site was at base A, the 202 bp DNA fragments were digested into two fragments, whose segment size was 127 and 75 bp respectively. When the restriction enzyme cutting site was at base C, the DNA fragments (202 bp) were digested into three fragments, with segment size of 105, 75 and 22 bp respectively. Among 300 samples, two kinds of alleles were detectable, which were A and C. There were three kinds of genotypes found in these 300 samples, exhibiting A/A wild type (127 bp, 75 bp), C/C homozygosis mutant (105, 75 and 22 bp) and A/C heterozygote mutant (127, 105, 75 and 22 bp) in genotypes (Figure S1). Among these 300 DNA samples, there were 276 cases of A/A wild type for CYP2C9*3 locus, which accounted for 92%. There were 22 cases of A/C heterozygote mutant, accounting for 7.3%. There were 2 cases of C/C homozygote mutant, accounting for 0.7%. The frequency of allele A was 95.67%, and that of allele C was 4.03%.

The DNA fragment of PCR and digesitive product at CYP2C9*3 locus was 22 bp. As it was too small, when

the gel electrophoresis finished, this fragment has run out of the gelatin, this fragment could not be seen within the galatin. Only DNA fragments sized 127, 105 and 75 bp could be observed.

The segment size of CYP2C9*c_65 locus derived from the PCR and digestion in 300 DNA samples was all 371 bp (Figure S3). By using restrictive nuclease *Hpa I*, when enzyme cutting site located at base G, 371 bp sized DNA fragment obtained from PCR was digested into two fragments, whose segment size was 317 and 54 bp respectively. When the enzyme cutting site located at base C, the DNA fragment of 371 bp could not digested by *Hpa I*, leaving a single fragment of 371 bp. Among the 300 DNA samples, there were two kinds of alleles detectable, which were G and C. There were two kinds of genotypes found in these 3000 DNA samples, exhibiting G/G wild type and G/C heterozygote mutant (Figure S2). Among these 300 DNA samples, there were 281 cases showing G/G wild type, accounted for 93.7%. There were 19 cases showing heterozygote mutant, accounting for 6.3%. The frequency of allele G was 96.83%, and that of allele C was 3.17% (Figure S3).

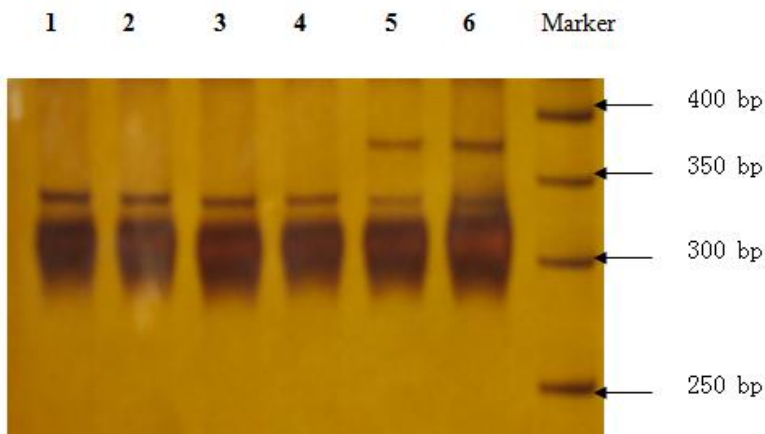


Figure S3. Showed genotypes of CYP2C9*c₆₅ locus. Number 1 to 4 were G/G of wild type (317 and 54 bp), Number 5-6 were heterozygote mutant (371, 317 and 54 bp).

Table 1. Hardy-Weinberg goodness of fit test of genotypes of CYP2C9*3 locus.

Frequency	A1/A1	A1/A2	A2/A2	χ^2	P
Actual frequency	276	22	2	4.002	0.177 (>0.05)
HW theoretical frequency	274.56	24.87	0.56		

Table 2. Hardy-Weinberg goodness of fit test of genotypes of CYP2C9*c₆₅ locus.

Frequency	A1/A1	A1/A2	A2/A2	χ^2	P
Actual frequency	281	19	0	0.321	0.451 (>0.05)
HW theoretical frequency	281.30	18.40	0.30		

Table 3. Sex distribution of CYP2C9*3 locus.

	CYP2C9*3 locus				CYP2C9*3 locus	
	cases	Genotype frequency			Allele frequency	
Gender		A1/A1	A1/A2	A2/A2	A	C
Male	106	94	10	2	198	14
Female	194	182	12	0	376	12
χ^2		0.581			0.134	
P		0.434 (>0.05)			0.714 (>0.05)	

DNA sequencing

Genetic analysis apparatus (3130 Genetic Analyzer, AB applied Biosystems) was used for sequencing the DNA fragments from PCR and restrictive nuclease digestion was conducted to confirm them as target DNA fragments. Results showed that sequencing outcome was in accordance with the standard DNA sequences of target DNA checked out from GeneBank.

Statistical analysis

Hardy-Weinbergbalance test (HWSIM) statistical analysis revealed that the observed number coincided well with the expected value in CYP2C9*3 and CYP2C9*c₆₅ locus (P>0.05), which was in accordance with the Hardy–Weinbergbalance law, suggesting that the DNA samples possessed group representativeness (Tables 1 and 2). Sex distribution of CYP2C9*3 and CYP2C9*c₆₅

Table 4. Sex distribution of CYP2C9*c₆₅ locus.

Gender	Genotype frequency			CYP2C9*c ₆₅ locus allele frequency		
	Cases	A1/A1	A1/A2	A2/A2	G	C
Male	106	101	5	0	207	5
Female	194	180	14	0	374	14
χ^2		1.127			0.602	
P		0.263 (>0.05)			0.405 (>0.05)	

DISCUSSION

In the present study, only the population of patients subjected to the operation of cardiac valve replacement was investigated. Because, in this population, patients routinely administered Warfarin, which provides an appropriate opportunity to examine the correlation of genotypes of special loci in CYP2C9 gene and Warfarin maintenance dosage. As for other diseases apart from cardiovascular disorders, such as diabetes, hypertension, were not involved in this study due to their independent with our study.

Influence of CYP2C9 gene mutation on Warfarin dosage in anticoagulant therapy and possible mechanism

The molecular mechanism of CYP2C9 gene polymorphism leading to metabolic defect of Warfarin lies on the base mutation, which results in alteration of sequences of DNA base, and then, amino acid replacement occurs, ultimately changes the catalytic activity of proteins, expressing as weak metabolic pattern and enzyme deficiency pattern. So far, researches involving in the correlation between CYP2C9 mutation and maintenance dosage of Warfarin in anticoagulant therapy demonstrated that CYP2C9 gene mutation could decrease the Warfarin metabolism. Therefore, patients with CYP2C9 gene mutation needed relatively lower dosage of Warfarin.

Higashi et al. (2002) firstly reported that the correlation between genotype of CYP2C9 gene and anticoagulation or hemorrhage. Subsequently, Sconce et al. (2005) found mutation of CYP2C9*2 or CYP2C9*3 reduced the Warfarin dosage needed in patients subjected to anticoagulant therapy, and CYP2C9*3 mutation led to a more large extent of Warfarin dosage reduction (30% reduced). There exists much variation of allele frequency in CYP2C9*2 and CYP2C9*3 loci among different races (Margaglione, 2000; Taube, 2000; Loebstein, 2001; Sanderson, 2005; Yu, 2004; Mizutani, 2003; Hong, 2005; Bae, 2005) as well. In White People, obvious ununiformity is commonly seen in the distribution of allele frequency of CYP2C9*2 gene in that the allele frequency

ranges from 8 to 19% (Nakai, 2005; Scordo, 2001; Garcia-Martin, 2001). Higashi et al. (2002) reported CYP2C9 gene has high genetic polymorphism, especially in the mutation of CYP2C9*2 and CYP2C9*3, because the activity of enzymes encoded by them decreased 30 and 80% respectively when compared with that of wild type CYP2C9*1, which is the main cause of CYP2C9 mutation, leading to the lower dosage of Warfarin administered in patients with CYP2C9 mutation. Although the correlation between CYP2C9*2 and/or CYP2C9*3 and Warfarin maintenance dose was involved in several studies by other authors, as for the correlation between CYP2C9*2 and/or CYP2C9*3 mutation and Warfarin maintenance dose in the Han people in China was few reported. Additionally, the crucial role of gene polymorphism of CYP2C9*2 and/or CYP2C9*3 in the intra-individual variation of Warfarin dose attracts more and more attention. Therefore, in this study, our finding that the influence of CYP2C9 gene polymorphism on the substrate drug metabolism has gene dosage effect sheds a new light on the personalized medication of Warfarin by applying Warfarin according to different genotypes of patients. Importantly, results from CYP2C9*3 locus study showed that the maintenance dose of Warfarin administered in anticoagulant therapy among the CYP2C9*3 genotypes exhibited: A/A wild type >A/C heterozygote >C/C homozygote, suggesting that patients carried with C/C mutation needed the lowest maintenance dose of Warfarin among the three genotypes found in this study. Our investigation is the first time to elucidate the conclusive relationship between the mutant in CYP2C9*3 gene locus and Warfarin maintenance dosage in Han people for clinical practice, providing a novel, useful and effective guidance for Warfarin personalized medication according to different genotypes in different patients.

Correlationship between CYP2C9*c₆₅ locus mutation and Warfarin maintenance dosage

In the present study, CYP2C9*c₆₅ locus-genotypes and their corresponding distributed rates suggesting the mutation at CYP2C9*c₆₅ locus may not correlated with maintenance dosage of Warfarin. However, Chen et al.

(2006) found in Chinese Taiwanese population that the mutation in CYP2C9*c₆₅ locus could reduce the Warfarin dosage administered, and in patients carried c₆₅ heterozygote or homozygote only needed one half of conventional dosage of Warfarin. To further ascertain the conclusive correlation between mutation of CYP2C9*c₆₅ locus and the Warfarin maintenance, we investigate whether or not this correlation exist in the Han people of China. The results of ours and other authors above mentioned were much different, it is indicative that CYP2C9*c₆₅ mutation has obvious variation among different populations and races. At least until now, there are no definite evidence revealing CYP2C9*c₆₅ mutation has conclusive relationship with Warfarin maintenance administered in anticoagulant therapy. This is another first report proposed by us from this study in Han people of China. We also found that there was no significant difference in genotype-distribution and allele frequency in CYP2C9*3 and CYP2C9 c₆₅ gene loci between male and female patients, suggesting CYP2C9 gene polymorphism had no definite correlation with gender variation.

Taken together, the maintenance dose of Warfarin administered in anticoagulant therapy among the CYP2C9*3 genotypes found in our experiment showed: A/A wild type> A/C heterozygote>C/C homozygote, suggesting that patients with C mutation need the lowest maintenance dose of Warfarin among the three genotypes found in this study. This will provide a conclusive and effective guidance for the Warfarin maintenance dose application according to different genotypes of patients. In future, intensive studies should focus on the correlation of more gene loci with Warfarin maintenance dose administered in the anticoagulant therapy and how to perform personalized medication of Warfarin in clinical practice.

ABBREVIATIONS

INR, International normalized ratio; **PT**, prothrombin time; **SNP**, single nucleotide polymorphism.

REFERENCES

- Aithal GP, Day CP, Kesteven P, Daly AK (1999). Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. *Lancet* 353:717-719.
- Alessandra P, Emma RT, Francis PS, David P, William HG, Sam M, Hibberd V, Holden J, Courtenay O, Wellington EM (2011). Pathogen Quantitation in Complex Matrices: A Multi-Operator Comparison of DNA Extraction Methods with a Novel Assessment of PCR Inhibition. *PLoS One* 6:e17916.
- Bae JW, Kim HK, Kim JH, Yang SI, Kim MJ, Jang CG, Park YS, Lee SY (2005). Allele and genotype frequencies of CYP2C9 in a Korean population. *Br. J. Clin. Pharmacol.* 60:418-422.
- Chern HD, Ueng TH, Fu YP, Cheng CW (2006). CYP2C9 polymorphism and warfarin sensitivity in Taiwan. *Chin. Chim. Acta.* 367:108-13.
- Garcia-Martin E, Martinez C, Ladero JM, Gamito FJ, Agúndez JA (2001). High frequency of mutations related to impaired CYP2C9 metabolism in a Caucasian population. *Eur. J. Clin. Pharmacol.* 57(1):47-49.
- Herman D, Locatelli I, Grabnar I, Peternel P, Stegnar M, Lainscak M, Mrhar A, Breskvar K, Dolzan V (2006). The influence of co-treatment with carbamazepine, amiodarone and statins on warfarin metabolism and maintenance dose. *Eur. J. Clin. Pharmacol.* 62:291-296.
- Higashi MK, Veenstra DL, Kondo LM, Wittkowsky AK, Srinouanprachanh SL, Farin FM, Rettie AE (2002). Association between CYP2C9 genetic variants and anticoagulation-related outcomes during warfarin therapy. *JAMA* 287:1690 - 1698.
- Higashi MK, Veenstra DL, Kondo LM, Wittkowsky AK, Srinouanprachanh SL, Farin FM, Rettie AE (2002). Association between CYP2C9 genetic variants and anticoagulation-related outcomes during warfarin therapy. *JAMA* 287: 1690-1698.
- Hong X, Zhang S, Mao G, Jiang S, Zhang Y, Yu Y, Tang G, Xing H, Xu X (2005). CYP2C9*3 allelic variant is associated with metabolism of irbesartan in Chinese population. *Eur. J. Clin. Pharmacol.* 61:627-634.
- Joanna J, Agnieszka M, Ryszard P (2012). Comparison of three methods of DNA extraction from human bones with different degrees of degradation. *Int. J. Legal Med.* 126:173-178.
- Lazarou J, Pomeranz BH, Corey PN (1998). Incidence of adverse drug reactions in hospitalized patients: a meta-analysis of prospective studies. *JAMA*, 279:1200-1205.
- Levine MN, Raskob G, Landefeld S, Kearon C (2001). Hemorrhagic complications of anticoagulant treatment. *Chest*, 119:108S-121S.
- Loebstein R, Yonath H, Peleg D, Almog S, Rotenberg M, Lubetsky A, Roitelman J, Harats D, Halkin H, Ezra D (2001). Inter-individual variability in sensitivity to warfarin: nature or nurture. *Clin. Pharmacol. Ther.* 70:159-164.
- Margaglione M, Colaizzo D, D'Andrea G, Brancaccio V, Ciampa A, Grandone E, Di Minno G (2000). Genetic modulation of oral anticoagulation with warfarin. *Thromb. Haemost.* 84:775-778.
- Mizutani T (2003). PM frequencies of major CYPs in Asians and Caucasians. *Drug Metab.* 35:99-106.
- Nakai K, Habano W, Nakai K, Fukushima N, Suwabe A, Moriya S, Osano K, Gurwitz D (2005). Ethnic differences in CYP2C9 * 2(Arg144Cys) and CYP2C9* 3(Ile359Leu) genotypes in Japanese and Israeli populations. *Life Sci.* 78: 107-111.
- Quick AJ (1935b). The prothrombin time in hemophilia and in obstructive jaundice. *J. Biol. Chem.* 109:73-74.
- Quick AJ, Stanley-Brown M, Bancroft FW (1935a). A study of the coagulation defect in hemophilia and in jaundice. *Am. J. Med. Sci.* 190:501-511.
- Sanderson S, Emery J, Higgins J (2005). CYP2C9 gene variants, drug dose, and bleeding risk in warfarin-treated patients: a Hu-Genet-systematic review and meta-analysis. *Genet. Med.* 7:97-104.
- Sconce EA, Khan TI, Wynne HA, Avery P, Monkhouse L, King BP, Wood P, Kesteven P, Daly AK, Kamali F (2005). The impact of CYP2C9 and VKORC1 genetic polymorphism and patient characteristics upon warfarin dose requirements: proposal for a new dosing regimen. *Blood* 106:2329-33.
- Scordo MG, Akillu E, Yasar U, Dahl ML, Spina E, Ingelman-Sundberg M (2001). Genetic polymorphism of cytochrome P450 2C9 in a Caucasian and a black African population. *Br. J. Clin. Pharmacol.* 52(4):447-450.
- Taube J, Halsall D, Baglin T (2000). Influence of cytochrome P-450 CYP2C9 polymorphisms on warfarin sensitivity and risk of over-anticoagulation in patients on long-term treatment. *Blood* 96:1816-1819.
- Yu BN, Luo CH, Wang D, Wang A, Li Z, Zhang W, Mo W, Zhou HH (2004). CYP2C9 allele variants in Chinese hypertension patients and healthy controls. *Clin. Chim. Acta* 348:57-61.

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